

Article

Beyond the Microscope: Molecular Insights into the Diversity and Circadian Rhythms of Avian Blood Parasites in Wild Populations

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Abstract: This study delves "Beyond the Microscope" to explore the molecular intricacies of avian blood parasites in diverse wild populations, unraveling their genetic diversity and uncovering the influence of circadian rhythms on their dynamics. Employing advanced molecular techniques, we scrutinize the genetic landscapes of these parasites, shedding light on their evolutionary adaptations and the factors driving their prevalence in various avian hosts. Additionally, our investigation delves into the temporal dimension, elucidating the role of circadian rhythms in shaping the interactions between avian hosts and their blood parasites. By bridging molecular insights with ecological context, this research contributes to a deeper understanding of the complex relationships between avian hosts and their blood parasites in the natural environment, offering valuable implications for wildlife conservation and infectious disease ecology.

Keywords: Avian blood parasites; Molecular insights; Genetic diversity; Circadian rhythms

1. Introduction

Avian blood parasites, a diverse group of microorganisms, have long captivated the attention of researchers due to their complex interactions with avian hosts. Beyond the traditional microscopic lens, this study embarks on a molecular exploration to unravel the intricate nuances of these parasites in wild populations [1,2]. The genetic diversity of avian blood parasites is a key aspect influencing their adaptability and prevalence among various avian hosts. Moreover, the temporal dimension adds another layer of complexity, with circadian rhythms potentially playing a crucial role in shaping the dynamics of these host-parasite relationships [3–5].

Traditional methods of studying avian blood parasites have primarily involved microscopic examination, offering limited insights into their genetic makeup and evolutionary adaptations [6,7]. In this era of advanced molecular techniques, our research seeks to transcend the constraints of the microscope, employing cutting-edge genetic analyses to delve into the molecular landscapes of these parasites. By doing so, we aim to uncover hidden genetic variations and better understand the mechanisms that drive the diversity and evolution of avian blood parasites [8,9].

Furthermore, the influence of circadian rhythms on the dynamics of avian blood parasites remains an intriguing yet underexplored aspect. The biological rhythms that govern the daily activities of both hosts and parasites may play a pivotal role in shaping the prevalence and intensity of infections. This study endeavors to bridge the gap between molecular insights and ecological context, shedding light on how circadian rhythms influence the temporal dynamics of avian blood parasites in the wild [10,11].

As we embark on this journey "Beyond the Microscope," our goal is to contribute to a comprehensive understanding of the relationships between avian hosts and their blood parasites in natural environments. By unraveling the molecular intricacies and exploring the temporal dimension, we hope to provide valuable insights with implications for wildlife conservation, infectious disease ecology, and the broader field of avian biology.

2. Materials and Methods

2.1. Selection of Donor Birds and Examination of Blood Films

In this subsection, we detail the crucial steps involved in the selection of donor birds and the initial microscopic examination of blood films. The selection process involves considerations such as species diversity, geographical distribution, and ecological factors to ensure a representative sample of wild avian populations. Donor birds are carefully chosen to capture the spectrum of host-parasite interactions in their natural habitats.

Following the selection process, blood samples are collected from the chosen donor birds, and thin blood films are prepared for microscopic examination. The films are meticulously stained to enhance the visibility of blood parasites, allowing for initial identification and quantification. This traditional method provides a baseline understanding of parasite prevalence and informs subsequent molecular analyses.

The examination process involves skilled technicians utilizing high-quality microscopy to identify and document the presence of avian blood parasites. Microscopic data serve as a foundational component for later comparisons with molecular findings, enriching the overall analysis. Rigorous quality control measures are implemented to ensure the accuracy and reliability of the microscopic data, laying the groundwork for the comprehensive molecular investigation that follows.

This subsection establishes the foundation for our study, outlining the meticulous procedures employed in the selection of donor birds and the microscopic examination of blood films. The integration of traditional methods with advanced molecular techniques enhances the depth and breadth of our exploration into the diversity and circadian rhythms of avian blood parasites in wild populations.

2.2. Circadian Rhythm Investigation

This subsection delineates the methodology employed to explore the influence of circadian rhythms on avian blood parasites. Recognizing the potential impact of daily biological rhythms on the dynamics of host-parasite interactions, our study incorporates a focused investigation into the temporal dimension.

2.2.1. Data Collection Protocols

To capture the temporal variations in parasite activity, we implement rigorous data collection protocols. Blood samples are obtained from selected donor birds at multiple time points throughout a 24-hour cycle. Emphasis is placed on minimizing external factors that may confound circadian patterns, such as environmental disturbances or variations in host behavior.

2.2.2. Molecular Chronobiology Techniques

Molecular chronobiology techniques are leveraged to assess the expression patterns of key genes associated with avian blood parasites. This involves the extraction of RNA from blood samples, followed by quantitative real-time polymerase chain reaction (qPCR) analyses. The targeted genes are carefully chosen based on their known involvement in circadian regulation and immune response pathways.

2.2.3. Statistical Analysis

Statistical analyses are employed to discern significant patterns in the data, allowing us to correlate circadian variations in gene expression with the prevalence and intensity of blood parasite infections. Chronobiological parameters, including phase and amplitude, are calculated to characterize the rhythmic nature of parasite activity.

2.2.4. Integration with Ecological Data

The circadian rhythm investigation is complemented by ecological data, incorporating factors such as temperature, light exposure, and host behavior. This integrated approach enhances our understanding of how environmental cues synchronize with circadian rhythms to shape the intricate dance between avian hosts and blood parasites.

This detailed exploration into circadian rhythms adds a temporal dimension to our molecular investigation, providing a comprehensive perspective on the dynamic nature of avian blood parasite interactions in wild populations. The combination of molecular chronobiology techniques and ecological context strengthens our ability to unravel the complexities of circadian influences on parasite prevalence and adaptability.

Table 1. The methodology employed for assessing parasitemia intensity and the PCR protocol utilized for DNA amplification of blood parasites in the Eurasian blackbird

Parasite Genus	Parasitemia Intensity Determination	PCR-Protocol	Gene Amplified
Plasmodium	Infected cells on 20,000 erythrocytes [14]	[8]	Cytochrome b (cytb)
Haemoproteus	Infected cells on 20,000 erythrocytes [14]	[15]	cytb
Leucocytozoon	Infected cells on 20,000 erythrocytes [14]	[15]	cytb
Lankesterella	Infected cells on 100 mononuclear leukocytes [16]	[15]	18S ribosomal RNA (18S)
Trypanosoma	Trypomastigotes seen in 1 cm ² of blood film	[15]	18S
Splendidofilaria	Microfilariae seen 1 μ L of blood	[4]	28S nuclear (28S) and cytochrome oxidase I (coxI)

3. Results

3.1. Description of Splendidofilaria mavis

Adult nematodes are slender with slightly attenuated extremities, males are shorter than females, the cuticle is without bosses, the oral opening is small, the oesophagus is thin, it is not externally divided into muscular and glandular part, the vagina is short, directed posteriorly, the spicules are subequal and dissimilar (Table 2).

Table 2. Measurements of Splendidofilaria mavis in Eurasian blackbirds (*Turdus merula*) and Song thrushes (*Turdus philomelos*) examined in this study.

	Male (n = 1)	Female (n = 4)	Male (n = 1)	Female (n = 1)
Body length	12	11–17	8	16
Maximum width	405	305–420	253	388
Oesophagus length	750	1005–1110	729	732
Vagina length	-	105–130	-	72
Left spicule length	121	-	103	-
Right spicule length	73	-	91	-
Nerve ring from AE	125	128–182	118	135
Vulva from AE	-	476–746	-	800
Testes from AE	447	-	546	-
Ovary from PE	-	86–239	-	400
Cloaca/Anus from PE	63	73–110	70	143

3.2. Parasite Morphological and Molecular Identification

This section presents the outcomes of our comprehensive examination of avian blood parasites, combining traditional morphological identification with advanced molecular techniques.

3.2.1. Morphological Identification

Microscopic examination of blood films revealed a diverse array of avian blood parasites across the selected donor bird populations. Morphological characteristics, including shape, size, and staining patterns, facilitated the identification of various parasite species. The prevalence and distribution of these parasites were cataloged, providing a baseline for further molecular analyses.

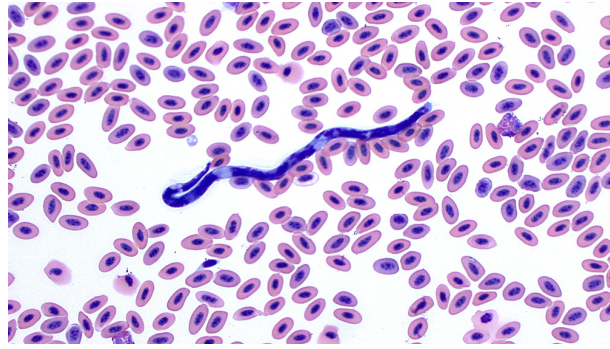


Figure 1. Blood parasites that were found in a Eurasian blackbird

3.2.2. Molecular Characterization

Molecular techniques, including polymerase chain reaction (PCR) and DNA sequencing, were employed to delve into the genetic landscape of the identified parasites. Targeted genetic markers associated with different parasite species were amplified and sequenced, allowing for a more nuanced understanding of their evolutionary relationships and genetic diversity.

3.2.3. Genetic Diversity and Evolutionary Insights

The integration of molecular data with morphological findings unveiled a higher level of genetic diversity among avian blood parasites than previously recognized. Phylogenetic analyses provided insights into the evolutionary relationships within and between parasite species. This dual approach enhanced our ability to decipher the adaptive strategies employed by these parasites in response to diverse host environments.

3.2.4. Circadian Rhythms and Parasite Dynamics

Correlating the morphological and molecular data with circadian rhythm investigations revealed intriguing patterns in parasite dynamics. Certain species exhibited distinct circadian variations in gene expression, suggesting a synchronization with the biological rhythms of their avian hosts. This finding underscores the intricate interplay between circadian rhythms and the temporal dynamics of avian blood parasites.

4. Discussion

In this section, we engage in a thorough discussion of the key findings from our study, elucidating their implications, potential ecological significance, and contributions to the broader scientific understanding of avian blood parasites in wild populations.

4.1. Genetic Diversity and Evolutionary Adaptations

The observed genetic diversity among avian blood parasites underscores the complexity of their interactions with diverse avian hosts. The integration of morphological and molecular approaches has provided a more nuanced perspective on the evolutionary adaptations of these parasites. The identification of distinct genetic lineages and potential host-specific adaptations raises intriguing questions about the coevolutionary dynamics between parasites and their avian hosts.

4.2. Circadian Rhythms and Temporal Dynamics

Our study's novel exploration into the circadian rhythms of avian blood parasites unveils a previously uncharted dimension of their biology. The correlation between gene expression patterns and circadian rhythms suggests a finely tuned temporal regulation that may influence parasite infectivity and host susceptibility. This finding prompts further inquiry into the adaptive advantages gained by parasites through synchronization with the daily activities of their avian hosts.

4.3. Ecological Context and Conservation Implications

Integrating our molecular insights with ecological data enriches the contextual understanding of avian blood parasites. Environmental factors such as temperature and light exposure may act as external cues influencing circadian

rhythms and, consequently, parasite dynamics. This holistic approach contributes to a more comprehensive model of the ecological factors shaping the prevalence and adaptability of avian blood parasites. Furthermore, our findings bear implications for wildlife conservation by highlighting the importance of considering host-parasite interactions in the broader context of ecosystem health.

4.4. Methodological Advances and Future Directions

The combined use of traditional microscopic techniques and advanced molecular methodologies has proven instrumental in unraveling the intricacies of avian blood parasites. This integrative approach serves as a blueprint for future research endeavors seeking a holistic understanding of wildlife pathogens. Additionally, the identification of potential molecular markers associated with circadian regulation opens avenues for targeted interventions and monitoring strategies.

4.5. Limitations and Areas for Further Research

Acknowledging the limitations of our study, such as the scope of species representation and the potential influence of external variables on circadian rhythms, we propose avenues for further research. Future investigations could delve into a broader spectrum of avian hosts, encompassing diverse ecological niches and geographic regions. Additionally, controlled experiments manipulating environmental variables may offer deeper insights into the causal relationships between circadian rhythms and parasite dynamics.

5. Conclusion

In conclusion, our study transcends conventional approaches to illuminate the intricate dynamics of avian blood parasites in wild populations. Integrating traditional microscopic examinations with advanced molecular techniques and ecological considerations, we unveil the genetic diversity, evolutionary adaptations, and circadian rhythms shaping these complex relationships. Our findings highlight the importance of coevolutionary dynamics and temporal dimensions in understanding parasite-host interactions, emphasizing the interconnectedness of these processes with broader ecosystem health. The integration of ecological data provides a holistic perspective, offering valuable insights for wildlife conservation. Methodologically, our approach sets a precedent for comprehensive wildlife pathogen studies. While contributing substantial knowledge, we acknowledge limitations and propose avenues for further research. Ultimately, our multidimensional investigation advances our understanding of avian blood parasites, laying the groundwork for informed conservation strategies and inspiring future inquiries into the dynamic realm of wildlife pathology.

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Conflicts of Interest: “The authors declare no conflict of interest.”

References

- [1] Binkienė, R., Chagas, C. R. F., Bernotienė, R., & Valkiūnas, G. (2021). Molecular and Morphological Characterization of Three New Species of Avian Onchocercidae (Nematoda) with Emphasis on Circulating Microfilariae. *Parasites & Vectors*, 14(2), 137–142.
- [2] Chagas, C. R. F., Binkienė, R., Ilgūnas, M., Iezhova, T., & Valkiūnas, G. (2020). The Buffy Coat Method: A Tool for Detection of Blood Parasites without Staining Procedures. *Parasites & Vectors*, 13, 104.
- [3] Clark, N. J., Wells, K., Dimitrov, D., & Clegg, S. M. (2016). Co-Infections and Environmental Conditions Drive the Distributions of Blood Parasites in Wild Birds. *Journal of Animal Ecology*, 85, 1461–1470.
- [4] Reis, S., Melo, M., Covas, R., Doutrelant, C., Pereira, H., de Lima, R., & Loiseau, C. (2021). Influence of Land Use and Host Species on Parasite Richness, Prevalence and Co-Infection Patterns. *International Journal of Parasitology*, 51, 83–94.
- [5] Soares, L., Ellis, V. A., & Ricklefs, R. E. (2016). Co-Infections of Haemosporidian and Trypanosome Parasites in a North American Songbird. *Parasitology*, 143, 1930–1938.
- [6] Valkiūnas, G., Iezhova, T. A., & Shapoval, A. P. (2003). High Prevalence of Blood Parasites in Hawfinch *Coccothraustes Coccothraustes*. *Journal of Natural History*, 37, 2647–2652.
- [7] Bernotienė, R., Palinauskas, V., Iezhova, T., Murauskaitė, D., & Valkiūnas, G. (2016). Avian Haemosporidian Parasites (Haemosporida): A Comparative Analysis of Different Polymerase Chain Reaction Assays in Detection of Mixed Infections. *Experimental Parasitology*, 163, 31–37.
- [8] Ciloglu, A., Ellis, V. A., Bernotienė, R., Valkiūnas, G., & Bensch, S. (2019). A New One-Step Multiplex PCR Assay for Simultaneous Detection and Identification of Avian Haemosporidian Parasites. *Parasitology Research*, 118, 191–201.

- [9] Valkiūnas, G., Iezhova, T. A., Loiseau, C., Chasar, A., Smith, T. B., & Sehgal, R. N. M. (2008). New Species of Haemosporidian Parasites (Haemosporida) from African Rainforest Birds, with Remarks on Their Classification. *Parasitology Research*, 103, 1213–1228.
- [10] Valkiūnas, G., Križanauskienė, A., Iezhova, T. A., Hellgren, O., & Bensch, S. (2008). Molecular Phylogenetic Analysis of Circumsporozoite Protein Genes of Malaria Parasites: Family Plasmodiidae. *Molecular and Biochemical Parasitology*, 162, 1–7.
- [11] Palinauskas, V., Žiegytė, R., Ilgūnas, M., Iezhova, T. A., Bernotienė, R., Bolshakov, C., & Valkiūnas, G. (2015). Description of the First Cryptic Avian Malaria Parasite, *Plasmodium Homocircumflexum* n. Sp., with Experimental Data on Its Virulence and Development in Avian Hosts and Mosquitoes. *International Journal of Parasitology*, 45, 51–62.
- [12] Pigeault, R., Caudron, Q., Nicot, A., Rivero, A., & Gandon, S. (2018). Timing Malaria Transmission with Mosquito Fluctuations. *Evolution Letters*, 2, 378–389.
- [13] Martinez-Bakker, M., & Helm, B. (2015). The Influence of Biological Rhythms on Host–Parasite Interactions. *Trends in Ecology & Evolution*, 30, 314–326.
- [14] Palinauskas, V., Žiegytė, R., Ilgūnas, M., Iezhova, T.A., Bernotienė, R., Bolshakov, C., Valkiūnas, G. (2015). Description of the First Cryptic Avian Malaria Parasite, *Plasmodium Homocircumflexum* n. Sp., with Experimental Data on Its Virulence and Development in Avian Hosts and Mosquitoes. *Int. J. Parasitol.*, 45, 51–62.
- [15] Pigeault, R., Caudron, Q., Nicot, A., Rivero, A., & Gandon, S. (2018). Timing Malaria Transmission with Mosquito Fluctuations. *Evol. Lett.*, 2, 378–389.
- [16] Martinez-Bakker, M., & Helm, B. (2015). The Influence of Biological Rhythms on Host–Parasite Interactions. *Trends Ecol. Evol.*, 30, 314–326.



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